REVIEW ARTICLE

Fats in Human Milk: 2022 Updates on Chemical Composition

Akhil Maheshwari

Received on: 15 November 2022; Accepted on: 25 November 2022; Published on: 23 December 2022

ABSTRACT

Human milk (HM) feedings are important for all newborn infants. Healthy term infants grow well with the mother's own milk (MOM), be it in direct breastfeeding or when fed expressed breastmilk. Premature and ill infants being treated/monitored in neonatal intensive care units (NICUs) also recover better when fed with HM diets, which can include MOM, donor milk (DM), or a combination of both. In terms of chemical composition, it contains 3–5% fat, 0.8–0.9% protein, 6.9–7.2% carbohydrates (calculated as lactose), and 0.2% mineral constituents. In this review, we present the latest information on HM fats, including triglycerides, phospholipids, triglycerides, cholesterol, glycoproteins, and enzymes. This article is intended to initiate a series of periodic updates on the scientific information available on HM fats. It contains some of our own research findings with an extensive review of the literature. To avoid bias in the identification of studies, keywords were short-listed *a priori* from anecdotal experience and from PubMed's Medical Subject Heading (MeSH) thesaurus. We then searched the databases PubMed, EMBASE, and Science Direct.

Keywords: Donor milk, Infants, Mother's own milk, Neonate, Neonatal intensive care unit, Newborn, Premature, Triglycerides. *Newborn* (2022): 10.5005/jp-journals-11002-0050

KEY POINTS

- Human milk feedings are important for all newborn infants. Healthy term infants grow well with MOM, be it in direct breastfeeding or when fed expressed breastmilk. Premature and ill infants being treated/monitored in NICUs recover better when fed with HM diets, which can include MOM, DM, or a combination of both.
- Fats in HM are an important source of energy for enterally fed growing term and premature infants. Human milk fats, including triglycerides, phospholipids, triglycerides, cholesterol, glycoproteins, and enzymes may facilitate recovery in ill neonates.
- Fats account for nearly 3–5% of HM and add up to nearly 25 grams/day during the first few months after birth. The amounts are sufficient to cover the physiological needs of newborn infants of about 4.8–6.6 grams/kg per day.
- Increasing information on various types of milk fats, including triacylglycerols, phospholipids, and saturated and unsaturated fatty acids, has improved our understanding of the roles of these lipids in development and disease.
- Docosahexaenoic acid (DHA, C22:6n-3) is a particularly important polyunsaturated FA. It is highly enriched in the brain and is being recognized for its role in neurodevelopment and retinal maturation.

INTRODUCTION

Human milk feedings are important for all newborn infants.^{1–3} Healthy term infants grow well with MOM, be it in direct breastfeeding or when fed expressed breastmilk.⁴ Premature and ill infants being treated/monitored in NICUs also recover better when fed with HM diets, which can include MOM, DM, or a combination of both.⁵ For these infants, milk may need appropriate fortification with bovine or HM-derived fortifiers.⁶ The use of donor HM, not formula, is also preferred to treat term infants with transient conditions such as hypoglycemia.⁷ Global Newborn Society, Clarksville, Maryland, United States of America

Corresponding Author: Akhil Maheshwari, Global Newborn Society, Clarksville, Maryland, United States of America, Phone: +1-7089108729, e-mail: akhil@globalnewbornsociety.org

How to cite this article: Maheshwari A. Fats in Human Milk: 2022 Updates on Chemical Composition. Newborn 2022;1(4):384–396.

Source of support: Nil Conflict of interest: None

The Joint Commission PC-05 and Baby-Friendly Hospital Initiative (BFHI) metrics include the exclusive use of HM throughout the initial newborn hospitalization.^{8,9} Human milk-derived fortifiers engineered from donated HM are also becoming popular in US NICUs, and are now used exclusively in 1/3rd or more of level-3–4 NICUs.¹⁰ The combination of HM-derived fortifiers with either DM or MOM, excluding any products cow's milk-containing products, has been labeled as exclusive HM diet.¹¹

Fat in HM is an important energy source for enterally fed growing term and premature infants.¹² Infants may have developmental limitations in utilizing milk-borne fats in the first few weeks after birth.¹³ The expression of many digestive enzymes and the transport of absorbed fats may be low.¹⁴ Maternal milk also shows changes in its fat composition during this period; the carbon chain length and the degree of unsaturation show maturational changes and are important determinants of absorption.¹⁵ The concentrations of major nutritional, immunological, and hormonal components in HM change over the course of lactation and differ between preterm and term populations.¹⁵ These components also vary based on the infant's birth order, gender, gestational age, and postnatal chronological age.¹⁶ Immunological factors are frequently influenced by infant's illness.¹⁶ Some HM hormones show diurnal variations in concentration.¹⁷ The long-term benefits of HM are promising for infants; there is a gut-lung and a gut-brain axis that

[©] The Author(s). 2022 Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (https://creativecommons. org/licenses/by-nc/4.0/), which permits unrestricted use, distribution, and non-commercial reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

links the intestinal microenvironment in early infancy with gut/lung maturation and neurodevelopmental outcomes.^{18,19}

In this review, we present the latest information on HM fats, including triglycerides, phospholipids, triglycerides, cholesterol, glycoproteins, and enzymes. All medical professionals involved in newborn care have long sought a comprehensive source of information on this very important topic, and after serial discussions in a subcommittee at the Global Newborn Society (GNS), we decided to collaborate across the organization to develop articles to cover various aspects of HM feedings ranging from clinical practice to its biochemistry. This article is intended to initiate a series of annual updates on the scientific information on HM. We searched extensively in the databases PubMed, EMBASE, and Scopus after short-listing keywords focused on the biochemistry and clinical relevance of these lipids.

Fats in HM

Fats (lipids) account for nearly 3–5% of HM, and are sufficient to cover the physiological needs of newborn infants of about 4.8–6.6 grams/kg per day.^{20,21} About 98% are triacylglycerols (TAGs) that carry nearly 88% of fatty acids (FAs).^{1,12} Small amounts of cholesterol esters (CEs) and phospholipids (PLs) are also seen.²² Table 1 summarizes these data. Overall, TAGs, FAs, CEs, and PLs comprise the four most important classes of lipids in HM and provide 50–60% of the total HM energy content.²³ Milk fats are important vehicles for the transport of many lipid-soluble hormones and vitamins A, D, E, and K.¹² Many of these components play important roles in the development of cognitive function and visual acuity.²⁴

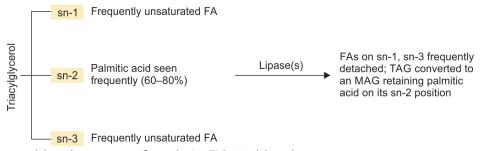
Fats make HM energy dense without an inappropriate increase in osmotic load.⁴ The highest concentrations are seen in colostrum and then decrease gradually in the postnatal period. This inverse correlation with rising milk volumes keeps the total daily ingested amounts of fats at a consistent level. The fat content shows considerable inter- and intra-individual variation.²⁵

Table 1: Types of fats in HM

Type of fats	Approximate percentages as constituents of the total fat
Triacylglycerols	98%
Diacylglycerols	Up to 1%
Monoacylglycerols	Traces
Nonesterified FAs	Up to 0.5%
Phospholipids	Up to 1%
Cholesterol	Up to 0.5%

The percentages are rounded-off figures from our own data

Flowchart 1: Schematic showing the structure of triacylglycerols detectable in HM



FA, fatty acid; MAG, monoacylglycerol; sn, stereospecific numbering; TAG, triacylglycerol

There is a rise in fat content each time a mother feeds her baby; hindmilk may contain up to 6% fats, which is more than the 2% seen in foremilk.²⁶ There is also a diurnal variation with higher fat contents during the day and lower during the night.¹⁷ All these variations contrast with the relatively constant concentrations of protein and lactose.

Triacylglycerols in HM

Triacylglycerols consist of a glycerol molecule that can be esterified in 3 FAs occupying segments, the sn (stereospecific numbering)-1, sn-2 (central position), and sn-3 (Flowchart 1).^{20,27} In this notation, the letter C and the following numerical modifier indicates the number of carbon atoms in the FA, and the numeral after the colon is the number of double bonds in the carbon chain.²⁸

Human milk contains at least 170 different types of TAGs, of which the 30 most abundant ones comprise 70% of the total milk-borne fats.^{29,30} In HM, 60–86% of the TAGs are esterified and these have a unique molecular structure compared to those seen in plasma and other tissues.³¹

Palmitic acid (hexadecanoic, C16:0) is the most frequently seen saturated FA in HM. It is usually located on the sn-2 position and accounts for 20–25% of the total FA content.^{32–34} This location is more conducive for absorption of lipids and calcium, bone health, intestinal flora, and overall comfort.^{35–37} Oleic acid (18:1 ω -9) is the most frequently seen unsaturated FA in HM.³³ Fatty acids are described by the ω - or the Δ -nomenclature; the ω names show the carbons counted from the methyl end, whereas the Δ nomenclature shows enumeration from the carboxylic acid.³⁸ Docosahexaenoic acid (cervonic; C22:3 ω -3) is also seen frequently, located on upto 50% of the sn-2 and 42% of the sn-3 positions.^{39–41} During digestion, pancreatic and gastric lipases may selectively hydrolyze FAs at the sn-1 and sn-3 positions, producing two free FAs and a 2-monoacylglycerol containing palmitic acid.⁴²

Monoacylglycerols containing palmitic acid are absorbed more efficiently, even more than free palmitic acid.³⁵ This is because palmitic acid has a melting point (61–65°C) that is higher than the body temperature, and because it forms insoluble soaps with calcium and magnesium in the intestine.^{43,44} In contrast, SFAs and 18-carbon FFAs such as oleic and linoleic acid are well-absorbed.⁴⁵ Details of various FAs seen in TAGs in HM are provided in the following sections:

Synthesis of TAGs Seen in HM

Mammary epithelial cells (MECs) contain large amounts of TAGs, most of which are synthesized locally.⁴⁶ Mammary epithelial cells contain the largest pool of FAs, nearly 95%, in the body⁴⁷; these are acquired via two processes:

Fatty acids synthesized within MECs from glucose in the endoplasmic reticulum (ER).⁴⁸ The rate-limiting step is the conversion of acetyl-CoA to malonyl-CoA, and the FA chains formed in this process grow via stepwise addition of 2-carbon units.⁴⁸ Fatty acid synthase is a key enzyme in this process.⁴⁹ Unlike in most other tissues where FA synthesis is terminated after a 16-carbon chain has been built, MECs contain an acyl thioester-hydrolase (thioesterase II) that can terminate FA synthesis at shorter lengths of 8–14 carbons to express mediumand intermediate-chain FAs.⁵⁰ These evolutionary adaptations can explain the high total FA content in milk.⁵¹

Alterations in maternal diet can change the FA composition of HM.⁵² Maternal diets low in fat and high in carbohydrates lead to *de novo* synthesis of FAs within the mammary gland, resulting in high concentrations of FAs of less than 16 carbons.⁴⁸ Therefore, although the total amount of fat present in the milk remains in the normal range, the fat is more saturated.

 Long-chain FAs (LCFAs; details in Table 1) imported from plasma, which are released from the digestion of TAGs (Flowchart 1) and are carried in circulating chylomicrons or very low-density lipoproteins (VLDLs).⁵³ These LCFAs in the bloodstream are bound to FA-binding proteins, or alternatively, combine first with CoA and then with an acyl-CoA-binding protein.^{54,55}

FAs formed in these two processes bind glycerol-3-phosphate to form TAGs, and these get incorporated into microlipid droplets.⁵⁶ These droplets coalesce and move outward toward the cell membrane, where those are pinched-off into the circulation.^{56,57}

Several technologies are now available to produce specifically tailored TAGs.⁵⁸ Biomimetic infant formulas containing TAG structures similar to those in HM may promote positive outcomes such as metabolic programming.⁵⁹ Lipid structures similar to those seen in HM can be produced *in vitro* by using specific lipases for interesterification.^{60,61} Several types of oils and fats, such as tripalmitin, lard fat, bovine milk fat, soybean oil, canola oil, borage oil, sunflower oil, and safflower oil, have been used to produce lipid structures resembling HM fat.^{62,63}

Digestion of HM-borne Saturated Fats

Neonates are developmentally deficient in many aspects of fat digestion because of low expression of lingual lipase, bile acids, and pancreatic lipase.⁶⁴ These enzymes may take a few days or more to be expressed at mature levels. Intragastric lipolysis by lingual and gastric lipases can partially compensate for the deficiency of pancreatic lipase.⁶⁴ Lingual lipase, secreted by the serous glands of the tongue, is detectable by 25 weeks' gestation.⁶⁵ Gastric lipase is secreted from the *chief (zymogen) cells* in the fundic gastric mucosa.⁶⁶ These can penetrate the milk lipid globules and hydrolyze the TAGs inside the core.⁶⁷ Fatty acids and monoglycerides resulting from intragastric lipolysis can compensate for low levels of bile acids by emulsifying lipid mixtures.⁶⁵

Human milk feedings can cover for many deficiencies in neonatal digestion as it carries many lipases, including lipoprotein lipase, bile salt esterase, and other nonactivated lipases.^{8,68,69} The composition of dietary fat should also be noted; the length of the carbon chain and the degree of unsaturation are important determinants of absorption.⁷⁰ Human milk supplies 8–12% of fat as medium-chain triglycerides (MCTs, chain length of 6–12 carbon atoms), which are hydrolyzed easier than long-chain triglycerides. In some cases, MCTs provide upto 40% of the total fat intake.⁷¹

Table 2: Phospholipids in human milk

	Mean (min–max) mg/100 g
Total phospholipids	25 (10–40)
Phosphatidylinositol	1 (1–2.5)
Phosphatidylserine	1.5 (1–2)
Phosphatidylethanolamine	7 (2–12)
Phosphatidylcholine	6 (2–10)
Sphingomyelin	9 (3–15)

Lipid digestion and absorption are also affected by dietary fat composition.⁷² Fatty acids with shorter chain length and higher degrees of unsaturation are absorbed more efficiently without the need for lipase or bile salts.⁷³ Therefore, commercial formulas are often designed to contain fats with more MCTs.⁷¹ These MCTs get digested and the FAs are transported directly to the liver via the portal vein. These MCTs can also enter mitochondria and be oxidized without the need for carnitine-mediated transport through mitochondrial membranes, and might even play a role in mitochondrial kinetics.^{74,75} However, further research is needed to understand the determinants of fat absorption or improved growth in preterm infants.

Phospholipids in HM

Human milk contains traces (<1%) of phospholipids, which interact with and form a protective membrane-like covering around bioactive components such as long-chain polyunsaturated FAs and choline.⁷⁶ We have provided some of our own data in Table 2 below.

Nonesterified Fatty Acids in HM

Fatty acids represent about 85% of the TAGs.³¹ As shown in Figure 1A, FAs are usually classified either by (a) the number of carbon atoms, into short-chain FAs (3–5 carbons in the longest chain), medium-chain (6–12 carbons), long-chain (13–22 carbons), and very-long-chain FAs (23–27 carbons);⁷⁷ or (b) the degree of saturation into saturated, monounsaturated, and polyunsaturated categories (Fig. 1).⁷⁸ Table 3 enlists various FAs in the three categories shown in Figure 1B.

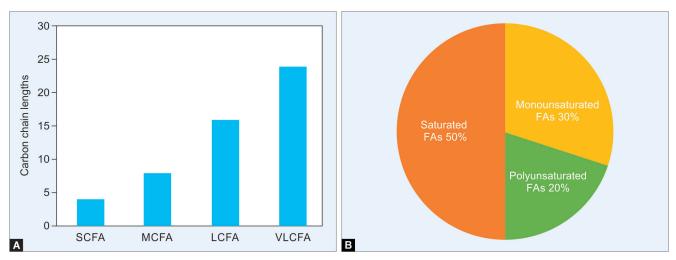
Saturated FAs in HM

About 45–50% of all FAs in HM are saturated, and these provide \geq 10–15% of the dietary energy content.⁷⁹ There are small amounts of short-chain (3–5 carbons long) saturated FAs and 8–10% are medium-chain (6–12 carbons long) FAs. Triacylglycerols containing these FAs are hydrolyzed by gut lipases without the need for bile salts, and the digested products are efficiently absorbed.⁸⁰

Besides known roles in nutrition, many saturated medium-chain FAs also perform important developmental functions.^{81,82} Caprylic (octanoic acid, C8:0) and capric (decanoic or decylic acid, C10:0) promote fat absorption.⁸³ Capric acid also exhibits bactericidal effects.⁸⁴ The LCFAs, including myristic (tetradecanoic, C14:0), palmitic (hexadecanoic, C16:0), arachidic (icosanoic acid, C20:0), and behenic (C22:0) acids, have immunomodulatory effects.⁷³ Palmitic acid may alter protein acylation.⁸⁵ It is also a key component of pulmonary surfactant.⁸⁶ In HM, palmitic acid is seen in more than half of all TAGs. Oleic acid is seen in 10–12%.¹

In HM, saturated FAs of carbon chain lengths 12, 14, 16, and 18 are seen in the highest concentrations.¹ The longer ones are a critical source of energy supply, and show dynamic plasticity.⁸⁷ Circulating stearic acid (octadecanoic, C18:0) is frequently interconverted





Figs 1A and B: (A) Structural classification of FAs based on lengths of the carbon chain (SCFA, short-chain FA; MCFA, medium-chain FA; LCFA, long-chain FA; VLCFA, very long-chain FA); (B) Classification of FAs based on saturation of carbon bonds in the fatty acid. The percentages show rounded-off data from our own laboratory

Saturated FAs		Mono-unsaturated FAs		Polyunsaturated FAs	
C8:0		C14:1 ω-5		C14:2 ω-6	
C10:0	Also called capric, decanoic, or decylic acid	C16:1 ω-9		C16:2 ω-6	
C12:0	Also called lauric, or dodecanoic acid	C16:1 ω-7	Also called palmitoleic acid	С18:2 ω-6	Also called linoleic acid
C13:0		C16:1 ω-5		cis, trans-C18:2 ω-6	
C14:0	Also called myristic, or tetradecanoic acid	C16:1 ω-3		CLA c9, t11	
C15:0		C17:1 ω-7		CLA c10, t11	
C16:0	Also called palmitic acid	C18:1 ω-9	Also called oleic acid	C18:3 ա-3	Also called α- linolenic acid
C17:0		C18:1 ω-7	A C18:1 (n-7) isomer of oleic acid. Also called cis-vaccenic or cis-11- octadecenoic acid.	C20:2 ω-6	
C18:0	Also called stearic acid	C18:1 ω-5		C20:3 ω-6	
C20:0		C20:1 ω-11		C20:4 ω-6	Also called arachidonic acid
C22:0		C20:1 ω-9		C22:2 ω-6	
C24:0		C22:1 ω-9		C22:4 ω-6	
				C22:5 ω-6	

Table 3: N	Aoct fr	oquantly	coon	EAcin	ЦИЛ
laple 3: r	viost fr	eauentiv	seen	FAS IN	HIM

Deep shading in the box indicates levels >10%, light shading indicates 1-10%, and no shading indicates <1%

with its monounsaturated counterpart, oleic acid.⁸⁸ Such transformations can be beneficial for preterm infants who may have limitations in absorbing fats. Although there is no demonstrated benefit for energy balance or growth in growing preterm infants, there is some evidence that many saturated medium-chain FAs can be beneficial in fat malabsorption due to short-bowel syndrome and severe cholestatic liver disease.⁷⁰

In humans, palmitic acid attached on the sn-2 position³⁵ renders it resistant to human lipases that can target sn-1,3 positions but not sn-2.⁸⁹ Consequently, lipase action leaves the TAGs only partially digested as sn-2 palmitate monoacylglycerols,^{37,90} which have these monoacylglycerols in digested HM, are better tolerated than free palmitic acid, augment fat and mineral absorption, and may also improve bone density in the medium term.^{31,37}

Litmanovitz et al.⁹¹ used ultrasound bone sonometry for longitudinal assessment of bone density in term and preterm infants. They performed a double-blinded, randomized controlled study of bone parameters in 3-month-old term infants fed formula containing triglycerides with *sn*-2 16:0 or standard vegetable oil blends and compared those to a nonrandomized group of breastfed infants. Infants in the intervention group had significantly higher bone density. These data were consistent with a previous report of Kennedy et al.,⁹² who had used dual-energy X-ray absorptiometry and shown higher bone mass in 3-month-old infants who were fed either formula containing structured triglycerides enriched in *sn*-2 16:0 or a conventional formula.

Palmitic acid in HM is important for lipoprotein and nonesterified FA metabolism when compared to that in TAGs sourced from plant oils.³¹ In the first few days after birth, HM contains lauric (dodecanoic, C12) and myristic (tetradecanoic acid, C14) saturated FAs in low concentrations.⁹³ As the concentrations of these two FAs rise, that of the longer FAs decreases.⁹⁴ Early FAs may also be derived from extramammary sources, but the breast then quickly begins to synthesize these.⁹⁵ The total fat content in milk may have a predictive value; 90% of the women whose milk contained \geq 20 grams fat per feeding on postnatal day 7 were successfully breastfeeding 3 months later.⁹⁶ Women with lower milk fat content had lower rates of success.⁹⁷

Mono- and Polyunsaturated Fatty Acids in HM

Monounsaturated FAs are an important component of HM and infant formulae.^{98,99} Oleic acid (OA, 18C:1 ω -9) is by far the most abundant monounsaturated FA.⁹⁹ In HM from Mediterranean regions, where consumption of olive oil is high, the concentrations of oleic acid and its congeners may exceed 40%, and the total content of monounsaturated FAs may exceed 45% of total FA in HM.^{100,101} These esterified FAs in TAGs interact extensively with the polyunsaturated FAs (PUFAs).¹⁰² We have described the interactions in the following sections:

The most frequently seen PUFAs in HM are listed in Table 1. These are derived from maternal diet, *de novo* synthesis in the mammary glands, and by mobilization from fat stores.³⁶ The FA composition of HM is influenced by many factors, including maternal diet, duration of pregnancy, maternal parity, and the stage of lactation.¹⁰³ Typically, the most abundant FAs are oleic (30%), palmitic (18%), linoleic (12%), myristic (6%), and stearic acids (8%).^{1,104,105} Interestingly, the concentrations of palmitoleic acid (an ω -7 monounsaturated FA, 16:1 ω -7) appear to mirror those in myristic acid.93 In preterm infants, HM may not always be adequate as a source of nutrients.8

Long-chain polyunsaturated FAs play an important role in the development of the infant's brain during the last trimester of pregnancy and during the first months after birth.¹⁰⁶ The precursor C18 fatty acids for the n-6 and n-3 LC-PUFAs are linoleic (C18:2 ω-6) and α -linolenic acid (C18:3 ω -3).¹⁰⁷ These are further elongated and desaturated to form other FAs, of which arachidonic acid (AA) and DHA are essential for normal growth and development.¹⁰⁸ Although the LC-PUFAs are synthesized from precursor FAs in both preterm and term infants, the capacity to produce DHA and AA is not known.¹⁰⁹ In the first week after birth, the levels of DHA and AA might drop due to the lack of adipose reserves and insufficient FA intake by the mother.¹¹⁰ The LC-PUFA content in HM in the United States, Europe, and Africa is similar, except for higher amounts of $\omega\text{-}3$ LC-PUFAs in the milk of women whose diets contain a large quantity of fish.^{111,112} Arachidonic acid (C20:4 ω-6) is an important LC-PUFA in HM.¹¹³ Eicosapentaenoic acid (EPA, C20:5 ω-3) is seen in relatively smaller quantities.^{113,114}

Polyunsaturated Essential FAs in HM

Human milk usually contains PUFAs such as linoleic acid (LA, 18:2 ω -6) and in adequate amounts.¹¹⁵ These FAs promote brain growth and retinal maturation, and influence metabolism such as by reducing plasma cholesterol.¹¹⁶ Human milk content of LA and

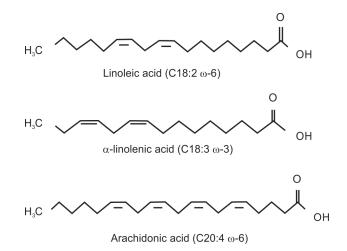


Fig. 2: The three essential FAs

alpha-linoleic acid (ALA) is known to vary according to maternal intake of these FAs.³⁶ The European Food Safety Authority (EFSA) recommends that infant formula should contain LA and ALA equivalent to 4.5–10.8% and 0.5–0.9% of the energy content.¹¹⁷ These are precursors of LC-PUFAs such as arachidonic acid and DHA.¹¹⁸

Linoleic acid, ALA, and ARA have been recognized as "essential fatty acids (EFAs)" as these cannot be synthesized endogenously and therefore, need to be acquired from diet in adequate amounts (Fig. 1).¹¹⁸ These FAs cannot be synthesized because the desaturases needed for introducing a double bond at carbons 3 and 6 (counted from the methyl end) are not expressed.⁸ Linoleic acid provides nearly 10% of the calories derived from the lipid fraction.⁹⁸ In preterm infants, biochemical evidence of EFA deficiency can be detected as early as 72 hours after birth.¹¹⁹

Long-chain Polyunsaturated FAs in HM

Increasing data now show that the capacity of young infants to synthesize LC-PUFAs might also be limited.¹²⁰ Hence, there may be a justification to supplement infant formulas with LC-PUFAs to bring the levels to resemble HM.¹²¹ The most frequently seen LC-PUFAs are shown in Figure 2. Maternal diet can alter the EFA and LC-PUFA content in HM. Several different elongases and desaturases transform EFAs to LC-PUFAs, such as arachidonic acid, DHA, and EPA, which then serve as substrates for bioactive metabolites such as eicosanoids, lipoxins, resolvins, and protectins.^{122,123} Fatty acids are also potential substrates for many elongases and desaturases for the formation of downstream metabolites.¹²⁴ In addition to these short-term changes, dietary changes can also change the lipids stored in her adipose tissues and consequently change the lipid composition of her milk.¹²⁵ Studies have shown a high correlation between the mother's fish intake and the DHA levels in her plasma and breast milk.⁴⁸ Therefore, maternal diet should receive special attention in lactating mothers.

There are some data that show a possible need for supplementing infant formula with monounsaturated FAs such as oleic (C18:1n-9) and palmitoleic acid (C16:1n-7, a product of palmitic acid metabolism).⁹⁸ These are normally secreted in HM in adequate amounts.³⁶ Oleic acid lowers the melting point of TAGs to enhance liquidity required for the formation, transport, and metabolism of milk fat.¹²⁶

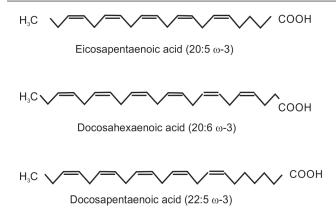


Fig. 3: The most frequently seen LC-PUFAs in human milk

Long-chain polyunsaturated FAs in HM may be derived from diet or drawn from FA storage pools in her own adipose tissue or the liver^{107,127} (Fig. 3). Dietary EFA and ω -3 FAs are also known to influence the LC-PUFA content in HM.³⁶ However, the effects of diet are relatively transient compared with the longer-term changes seen with FAs drawn from body's internal pools.¹²⁸ Fatty acids that are synthesized, elongated, or desaturated in the liver or peripheral tissues are more likely to be incorporated into milk fat.¹²⁸ Most studies on these changes have focused on the linoleic acid, α -linolenic acid EFAs. A review of 65 studies showed lower variation of ARA compared with DHA between populations with different dietary habits.¹²⁹

The perinatal period is a period of intensive growth of the brain, which contains large amounts of ARA and DHA.¹³⁰ There is a likely need for LC-PUFAs in larger amounts for the development of neurological and cognitive functions.¹³¹ Long-chain polyunsaturated FAs are also involved in inflammatory and immunological processes, suggesting a need in optimal maturation of the immune system.^{132–134} Long-chain polyunsaturated FAs might also influence adipocyte differentiation and consequently, the risk of obesity in childhood or later.¹²⁰

Carnitine promotes the transport of LC-PUFAs into the mitochondria for oxidation and removal of short-chain FAs that accumulate in mitochondria.¹³⁵ Preterm infants are at risk for relative deficiency of carnitine because of their dependence on lipids as an energy source and limited endogenous synthetic ability, although the clinical need is still unproven.¹³⁶ Carnitine is detectable in HM and is currently added to standard term and preterm formulas.¹³⁷ In preterm infants not receiving supplemental carnitine, plasma and tissue carnitine levels may fall even in the presence of adequate precursor amino acid concentrations.^{136,138}

Yaron et al.¹³⁹ showed that infants fed with a formula supplemented with β -16:0 for 6 weeks showed more *Lactobacillus* and *Bifidobacterium* genera in their stools than a control group that received formula containing vegetable-sourced 16:0. The mechanisms are unclear, but there is a possibility that the position of palmitic acid on the TAGs may have influenced that gut microbiome.¹⁴⁰ Current infant formulas typically include palm, coconut, soybean, and sunflower oils as the primary lipid sources.⁹⁸ The development of structured lipids that resemble HM fat may improve the safety of formula feedings.⁷³

The synthesis for LC-PUFAs in HM has severe unique patterns. Similar to the desaturases and elongases seen in plasma, red blood cells, and the adipose tissue, the efficacy of similar enzymes seen in HM in conversion of EFAs into LC-PUFAs may determine the relative concentrations of various FAs in milk.¹³² However, there may be subtle differences between individual FAs. Alphalinolenic acid, the precursor of the n-3 series LC-PUFAs, is processed differently than LA.³⁶ Nearly 65% of milk ALA seems to be directly derived from maternal diet, not from the body stores.^{36,141} The association of maternal DHA intake (fish oil or other sources of ω -3 LC-PUFAs) with milk DHA levels is stronger than that of maternal ARA intake.¹⁴² There are a few studies that tested the effects of n-3 supplementation on milk fat composition in combination with ARA supplementation.³⁶ In a Dutch study, 88 breastfeeding women received DHA (220 mg/day), with milk samples collected in the second and 12th week of lactation.^{143,144} Docosahexaenoic acid percentages were significantly higher than in the placebo group. In the supplemented group, the addition of ARA led to higher median ARA percentages in milk. As the supplementation had started already mid-pregnancy, the findings suggested that increased ARA excretion with milk could be the result of increased direct transfer from the diet and increased ARA contribution to the flux of fatty acids into milk passing maternal storage pools. Several other studies have shown similar findings.^{145–147} There is a need to re-evaluate current beliefs that only 10% of milk ARA is directly derived from the diet.

DHA in HM

Docosahexaenoic acid is an important LC-PUFA in HM, most of it *is* derived from its precursor, α -linolenic acid (18:3 ω -3).^{148,149} It is highly enriched in the brain and is important for neurodevelopment.¹³¹ Infants, particularly those born preterm, can develop LC-PUFA deficiencies because of their limited ability to synthesize these FAs.¹¹⁰ Here, an important adaptation in the mother–infant axis is that preterm milk contains more C8–C14 FAs and LC-PUFAs than term milk, and the LC-PUFA content gradually decreases with increasing postnatal age.¹¹⁵ These developmental adaptations may be helpful as shorter FAs might be easier to digest, and LC-PUFAs are important for brain and retinal development.⁸⁰

Docosahexaenoic acid is important for neurologic development and advancement of visual acuity.¹¹⁶ Lipids are an important constituent of brain matter, and DHA constitutes 30–40% of the fatty acids in the gray matter and the synaptic membranes.¹⁵⁰ The retina, particularly its outer rod segment, also has a high DHA content.¹⁵¹ Fatty acids are known to alter gene expression in the developing brain, and hence, are likely to alter long-term neurodevelopment and metabolism.¹⁵² These effects are most likely to be seen in the fetus/premature infants, who may not yet have the capacity to absorb/synthesize specific FAs such as the LC-PUFAs.¹⁵³

Western diets may not always provide appropriate amounts of DHA for lactating mothers, and DHA levels may fall by 40% in preterm infants during the first week after birth.¹⁵⁴ Many studies have measured the effects of DHA supplementation in lactating mothers on the FA composition in HM and infant plasma.^{113,155–157} Maternal daily supplementation of 1200 mg of DHA can increase HM and infant plasma DHA concentrations to almost 12 and 2–3 times higher than in controls.¹⁵⁸ These higher levels remained elevated even after 6 weeks of maternal supplementation of DHA at 400 mg/ day.^{159,160} Current evidence suggests that DHA supplementation to lactating mothers is safe and effective in increasing DHA levels in HM.¹⁶¹ Some infant formulas are now being supplemented with these FAs.^{162,163} In preterm infants, raising the DHA levels by 2–3 times may have improved neurodevelopmental and cognitive outcomes such as information processing.^{164,165} In term infants, LC-PUFA supplementation may improve visual acuity by 12 months of age.¹⁶⁶ Cholesterol is another small (9–12 mg/dL), but an important lipid constituent of HM.¹⁶⁷ Breastfed infants may receive more cholesterol per kg body weight in feedings than adults but have better lipid profiles.¹⁶⁸ There is a possibility that early breastfeeding associated with high measured total blood serum cholesterol may prevent, not raise, some of the risks of developing cardiovascular diseases later in life.¹⁶⁹

In preterm infants, DHA and its downstream metabolites, the oxylipins, are important regulators of inflammatory responses.¹⁷⁰ The deficiency of DHA after birth can augment inflammation, particularly in preterm infants.^{171,172} Infants with higher mean DHA levels may be less likely to develop chronic lung disease (CLD).¹⁷³ Docosahexaenoic acid supplementation may also be protective against necrotizing enterocolitis (NEC).¹⁷⁴ The mechanisms by which DHA supplementation to lactating mothers or to infants attenuates inflammation are still not clear; one possibility is that N-docosahexaenoylethanolamine (synaptamide), a neurogenic and synaptogenic metabolite of DHA, may mediate some of these anti-inflammatory effects.^{175,176} Maternal diet high in ω -3 fatty acids upregulates genes involved in neurotrophin signaling in fetal brain during pregnancy in C57BL/6 mice.¹⁷⁷ The mechanisms of DHA effects in humans are not clear.

Docosahexaenoic acid and other LC-PUFAs can be acquired in diet, but humans can also synthesize some small quantities from precursors linoleic acid (18:2 ω -6) and α -linolenic acid (18:3 ω -3).¹⁰⁷ The most important pathways include Δ 4-desaturation, β -oxidation, and carbon recycling.¹⁷⁸ The FA desaturase (FADS) gene cluster consists of a family of three genes located on human chromosome 11q12-13.1 that yields enzymes catalyzing the insertion of double bonds in PUFA, monounsaturated FAs and palmitic acid.¹⁷⁹ FADS2 can code for pathways involving Δ 6-, Δ 8-, and Δ 4-desaturation.¹⁸⁰ FADS1 codes for a Δ 5-desaturase, leading directly to the signaling precursor ω -6 arachidonic acid (20:4 ω -6) and to ω -3 eicosapentaenoic acid (20:5 ω -3).^{162,181} Despite these well-established classical results, substrate competition is known to modulate the relative activity of desaturases that defines total PUFA composition of tissues.¹⁸²

In parenteral nutrition, many lipid preparations now contain DHA and AA.^{183,184} However, these preparations may not always correct postnatal FA deficits.¹⁸⁵ Eicosapentaenoic acid levels may be elevated due to the fish oil component in these lipid preparations but the DHA and AA levels may decline.^{186–190} Low AA levels could possibly be associated with suboptimal clinical outcomes such as with increased risk of late-onset sepsis and retinopathy of prematurity.^{171,191,192}

Docosahexaenoic acid and AA are naturally expressed in HM, and so most infant formulas for term and preterm infants are now supplemented with these FAs.¹⁶³ Many RCTs have been conducted to evaluate the addition of DHA and AA to preterm formulas.^{109,193} Most studies show positive or no changes in growth, although few show negative effects.¹⁹³ Findings of improved visual acuity have been inconsistent. Formula supplemented with DHA and AA seems to improve visual acuity,¹⁶⁶ but the effects on neurodevelopment remain unclear.¹⁹³ Current recommendations advocate for VLBW infants to receive 55–60 mg of DHA and 35–45 mg per kg/day of AA.^{194,195}

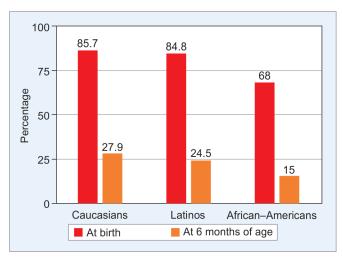


Fig. 4: HM feeding rates by race in the United States (CDC, 2017)

A recent Cochrane review¹⁹⁶ showed that ω -3 LC-PUFA supplementation during pregnancy is effective in reducing the incidence of preterm birth, although it may increase post-term pregnancies. Preterm birth <37 weeks and early preterm birth <34 weeks were reduced in women receiving omega-3 LCPUFA compared with no ω -3.¹⁹⁶ There as a possibility of reduced risk of perinatal death and of neonatal care admission, reduced risk of LBW babies; but a small increased risk of LGA babies with ω -3 LC-PUFAs.¹⁹⁶ For GRADE quality assessments,¹⁹⁷ the conclusions for perinatal outcomes were viewed as high- or moderate-quality evidence.

CONCLUSION

Human milk is the primary source of nutrients for neonates, and may be the best option from the points of view of nutritional, immunological, food safety, and growth and development.¹⁹⁸ It also improves mother–infant bonding, and facilitates the emotional, cognitive, and nervous system development of the infant.¹⁹⁹ Unfortunately, rapid urbanization and social limitations have been major constraints in promoting HM feedings.^{200,201} Worldwide, only 35–40% of infants receive HM from birth to 6 months of age.^{202–205} Breastfeeding rates diverge widely along the lines of race, socioeconomic status, and ethnicity (summarized in Figure 4).^{206,207}

Despite all the educational and logistical strategies to promote HM feeding, some infants will still continue to need at least some formula feedings because of medical, social, or other reasons.²⁰⁸ Therefore, there will be a need to develop and improve infant formulas that closely mimic the nutritional and chemical characteristics of HM.²⁰⁹ Timely delivery of physiologically important lipids will be needed to improve growth and development by optimizing the energy contents, without unduly increasing the osmolar loads.^{73,210} We also cannot overemphasize the importance of lipids in neurological development, protection of the gastrointestinal tract, immune defenses, and cholesterol metabolism.^{52,154,211,212}

REFERENCES

- Jensen RG, Hagerty MM, McMahon KE. Lipids of human milk and infant formulas: A review. Am J Clin Nutr 1978;31(6):990–1016. DOI: 10.1093/ajcn/31.6.990.
- Jenness R. The composition of human milk. Semin Perinatol 1979; 3(3):225–239. PMID: 392766.

- Valverde R, Dinerstein NA, Vain N. Mother's own milk and donor milk. World Rev Nutr Diet 2021;122:212–224. DOI: 10.1159/000514733.
- Martin CR, Ling PR, Blackburn GL. Review of infant feeding: Key features of breast milk and infant formula. Nutrients 2016;8(5):279. DOI: 10.3390/nu8050279.
- Villamor-Martinez E, Pierro M, Cavallaro G, et al. Mother's own milk and bronchopulmonary dysplasia: A systematic review and metaanalysis. Front Pediatr 2019;7:224. DOI: 10.3389/fped.2019.00224.
- 6. Arslanoglu S, Boquien CY, King C, et al. Fortification of human milk for preterm infants: Update and recommendations of the European Milk Bank Association (EMBA) Working Group on human milk fortification. Front Pediatr 2019;7:76. DOI: 10.3389/fped.2019.00076.
- 7. Ferrarello D, Schumacher A, Anca R. Nurse-driven initiative to increase exclusive human milk feeding by using pasteurized donor human milk to treat hypoglycemic term neonates. Nurs Womens Health 2019;23(4):316–326. DOI: 10.1016/j.nwh.2019.05.001.
- (v2015B) SMfJCNQM. Exclusive breast milk feeding during the newborn's entire hospitalization. https://manual.jointcommission. org/releases/TJC2015B/MIF0170.html.
- 9. Fair FJ, Morrison A, Soltani H. The impact of baby friendly initiative accreditation: An overview of systematic reviews. Matern Child Nutr 2021;17(4):e13216. DOI: 10.1111/mcn.13216.
- Premkumar MH, Pammi M, Suresh G. Human milk-derived fortifier versus bovine milk-derived fortifier for prevention of mortality and morbidity in preterm neonates. Cochrane Database Syst Rev 7 2019;2019(11):CD013145. DOI: 10.1002/14651858.CD013145.pub2.
- Bushati C, Chan B, Harmeson Owen A, et al. Challenges in implementing exclusive human milk diet to extremely low-birthweight infants in a level III neonatal intensive care unit. Nutr Clin Pract 2021;36(6):1198–1206. DOI: 10.1002/ncp.10625.
- 12. Koletzko B. Human milk lipids. Ann Nutr Metab 2016;69(Suppl 2): 28–40. DOI: 10.1159/000452819.
- Amissah EA, Brown J, Harding JE. Fat supplementation of human milk for promoting growth in preterm infants. Cochrane Database Syst Rev 2020;2020(8):CD000341. DOI: 10.1002/14651858.CD000341. pub3.
- 14. Manson WG, Weaver LT. Fat digestion in the neonate. Arch Dis Child Fetal Neonatal Ed 1997;76(3):F206–F211. DOI: 10.1136/fn.76.3.f206.
- Ballard O, Morrow AL. Human milk composition: Nutrients and bioactive factors. Pediatr Clin North Am 2013;60(1):49–74. DOI: 10.1016/ j.pcl.2012.10.002.
- Nolan LS, Parks OB, Good M. A review of the immunomodulating components of maternal breast milk and protection against necrotizing enterocolitis. Nutrients 2019;12(1):14. DOI: 10.3390/nu1201 0014.
- 17. Italianer MF, Naninck EFG, Roelants JA, et al. Circadian variation in human milk composition, a systematic review. Nutrients 2020; 12(8):2328. DOI: 10.3390/nu12082328.
- Rodriguez JM, Fernandez L, Verhasselt V. The GutBreast axis: Programming health for life. Nutrients 2021;13(2):606. DOI: 10.3390/ nu13020606.
- 19. Ratsika A, Codagnone MC, O'Mahony S, et al. Priming for life: Early life nutrition and the microbiota-gut-brain axis. Nutrients 2021;13(2):423. DOI: 10.3390/nu13020423.
- Ahmadian M, Duncan RE, Jaworski K, et al. Triacylglycerol metabolism in adipose tissue. Future Lipidol 2007;2(2):229–237. DOI: 10.2217/17460875.2.2.229.
- Eibensteiner F, Auer-Hackenberg L, Jilma B, et al. Growth, feeding tolerance and metabolism in extreme preterm infants under an exclusive human milk diet. Nutrients 2019;11(7):1443. DOI: 10.3390/ nu11071443.
- Emken EA, Adlof RO, Hachey DL, et al. Incorporation of deuteriumlabeled fatty acids into human milk, plasma, and lipoprotein phospholipids and cholesteryl esters. J Lipid Res 1989;30(3):395–402. PMID: 2723546.
- 23. George AD, Gay MCL, Trengove RD, et al. Human milk lipidomics: Current techniques and methodologies. Nutrients 2018;10(9):1169. DOI: 10.3390/nu10091169.

- Lee H, Park H, Ha E, et al. Effect of breastfeeding duration on cognitive development in infants: 3-year follow-up study. J Korean Med Sci 2016;31(4):579–584. DOI: 10.3346/jkms.2016.31.4.579.
- 25. Selvalatchmanan J, Rukmini AV, Ji S, et al. Variability of lipids in human milk. Metabolites 2021;11(2):104. DOI: 10.3390/metabo11020104.
- Mizuno K, Nishida Y, Taki M, et al. Is increased fat content of hindmilk due to the size or the number of milk fat globules? Int Breastfeed J 2009;4:7. DOI: 10.1186/1746-4358-4-7.
- 27. Karupaiah T, Sundram K. Effects of stereospecific positioning of fatty acids in triacylglycerol structures in native and randomized fats: A review of their nutritional implications. Nutr Metab (Lond) 2007;4:16. DOI: 10.1186/1743-7075-4-16.
- McConathy J, Owens MJ. Stereochemistry in drug action. Prim Care Companion J Clin Psychiatry 2003;5(2):70–73. DOI: 10.4088/pcc. v05n0202.
- Yuan T, Qi C, Dai X, et al. Triacylglycerol composition of breast milk during different lactation stages. J Agric Food Chem 2019;67(8): 2272–2278. DOI: 10.1021/acs.jafc.8b06554.
- Zhu H, Liang A, Wang X, et al. Comparative analysis of triglycerides from different regions and mature lactation periods in Chinese Human Milk Project (CHMP) study. Front Nutr 2021;8:798821. DOI: 10.3389/fnut.2021.798821.
- 31. Innis SM. Dietary triacylglycerol structure and its role in infant nutrition. Adv Nutr 2011;2(3):275–283. DOI: 10.3945/an.111.000448.
- Straarup EM, Lauritzen L, Faerk J, et al. The stereospecific triacylglycerol structures and fatty acid profiles of human milk and infant formulas. J Pediatr Gastroenterol Nutr 2006;42(3):293–299. DOI: 10.1097/01.mpg.0000214155.51036.4f.
- Jensen RG. Lipids in human milk. Lipids 1999;34(12):1243–1271. DOI: 10.1007/s11745-999-0477-2.
- Breckenridge WC, Marai L, Kuksis A. Triglyceride structure of human milk fat. Can J Biochem 1969;47(8):761–769. DOI: 10.1139/o69-118.
- Carta G, Murru E, Banni S, et al. Palmitic acid: Physiological role, metabolism and nutritional implications. Front Physiol 2017;8:902. DOI: 10.3389/fphys.2017.00902.
- Giuffrida F, Fleith M, Goyer A, et al. Human milk fatty acid composition and its association with maternal blood and adipose tissue fatty acid content in a cohort of women from Europe. Eur J Nutr 2022;61(4): 2167–2182. DOI: 10.1007/s00394-021-02788-6.
- 37. Bar-Yoseph F, Lifshitz Y, Cohen T. Review of sn-2 palmitate oil implications for infant health. Prostaglandins Leukot Essent Fatty Acids 2013;89(4):139–143. DOI: 10.1016/j.plefa.2013.03.002.
- Davidson BC, Cantrill RC. Fatty acid nomenclature. A short review. S Afr Med J 1985;67(16):633–634. DOI: 10.1002/chin.198542383.
- Hunter JE. Studies on effects of dietary fatty acids as related to their position on triglycerides. Lipids 2001;36(7):655–668. DOI: 10.1007/ s11745-001-0770-0.
- Valenzuela A, Nieto S, Sanhueza J, et al. Tissue accretion and milk content of docosahexaenoic acid in female rats after supplementation with different docosahexaenoic acid sources. Ann Nutr Metab 2005;49(5):325–332. DOI: 10.1159/000087337.
- Wu K, Gao R, Tian F, et al. Fatty acid positional distribution (sn-2 fatty acids) and phospholipid composition in Chinese breast milk from colostrum to mature stage. Br J Nutr 2019;121(1):65–73. DOI: 10.1017/ S0007114518002994.
- 42. Lien EL. The role of fatty acid composition and positional distribution in fat absorption in infants. J Pediatr 1994;125(5 Pt 2):S62–S68. DOI: 10.1016/s0022-3476(06)80738-9.
- van Rooijen MA, Mensink RP. Palmitic acid versus stearic acid: Effects of interesterification and intakes on cardiometabolic risk markers – A systematic review. Nutrients 2020;12(3):615. DOI: 10.3390/nu12030615.
- 44. Graham DY, Sackman JW. Solubility of calcium soaps of longchain fatty acids in simulated intestinal environment. Dig Dis Sci 1983;28(8):733–736. DOI: 10.1007/BF01312564.
- Young RJ, Garrett RL. Effect of oleic and linoleic acids on the absorption of saturated fatty acids in the chick. J Nutr 1963;81(4): 321–329. DOI: 10.1093/jn/81.4.321.

- 46. Yonezawa T, Yonekura S, Kobayashi Y, et al. Effects of long-chain fatty acids on cytosolic triacylglycerol accumulation and lipid droplet formation in primary cultured bovine mammary epithelial cells. J Dairy Sci 2004;87(8):2527–2534. DOI: 10.3168/jds.S0022-0302(04)73377-9.
- 47. McManaman JL. Formation of milk lipids: A molecular perspective. Clin Lipidol 2009;4(3):391–401. DOI: 10.2217/clp.09.15.
- Aumeistere L, Ciprovica I, Zavadska D, et al. Impact of maternal diet on human milk composition among lactating women in Latvia. Medicina (Kaunas) 2019;55(5):173. DOI: 10.3390/medicina 55050173.
- Beld J, Lee DJ, Burkart MD. Fatty acid biosynthesis revisited: Structure elucidation and metabolic engineering. Mol Biosyst 2015;11(1):38–59. DOI: 10.1039/c4mb00443d.
- Ritchie MK, Johnson LC, Clodfelter JE, et al. Crystal structure and substrate specificity of human thioesterase 2: Insights into the molecular basis for the modulation of fatty acid synthase. J Biol Chem 2016;291(7):3520–3530. DOI: 10.1074/jbc.M115.702597.
- 51. Randhawa ZI, Naggert J, Blacher RW, et al. Amino acid sequence of the serine active-site region of the medium-chain S-acyl fatty acid synthetase thioester hydrolase from rat mammary gland. Eur J Biochem 1987;162(3):577–581. DOI: 10.1111/j.1432-1033.1987. tb10678.x.
- Innis SM. Impact of maternal diet on human milk composition and neurological development of infants. Am J Clin Nutr 2014;99(3): 734S–741S. DOI: 10.3945/ajcn.113.072595.
- Sundaram M, Yao Z. Recent progress in understanding protein and lipid factors affecting hepatic VLDL assembly and secretion. Nutr Metab (Lond) 2010;7:35. DOI: 10.1186/1743-7075-7-35.
- Furuhashi M, Hotamisligil GS. Fatty acid-binding proteins: Role in metabolic diseases and potential as drug targets. Nat Rev Drug Discov 2008;7(6):489–503. DOI: 10.1038/nrd2589.
- Haunerland NH, Spener F. Fatty acid-binding proteins--insights from genetic manipulations. Prog Lipid Res 2004;43(4):328–349. DOI: 10.1016/j.plipres.2004.05.001.
- 56. Gao Q, Goodman JM. The lipid droplet-a well-connected organelle. Front Cell Dev Biol 2015;3:49. DOI: 10.3389/fcell.2015.00049.
- Olzmann JA, Carvalho P. Dynamics and functions of lipid droplets. Nat Rev Mol Cell Biol 2019;20(3):137–155. DOI: 10.1038/s41580-018-0085-z.
- Farfan M, Alvarez A, Garate A, et al. Comparison of chemical and enzymatic interesterification of fully hydrogenated soybean oil and walnut oil to produce a fat base with adequate nutritional and physical characteristics. Food Technol Biotechnol 2015;53(3):361–366. DOI: 10.17113/ftb.53.03.15.3854.
- Bourlieu C, Mahdoueni W, Paboeuf G, et al. Physico-chemical behaviors of human and bovine milk membrane extracts and their influence on gastric lipase adsorption. Biochimie 2020;169:95–105. DOI: 10.1016/j.biochi.2019.12.003.
- 60. Xie W, Qi C. Interesterification of soybean oil and lard blends catalyzed by SBA-15-pr-NR(3)OH as a heterogeneous base catalyst. J Agric Food Chem 2013;61(14):3373–3381. DOI: 10.1021/jf400216z.
- 61. Xie W, Zang X. Immobilized lipase on core-shell structured Fe3O4-MCM-41 nanocomposites as a magnetically recyclable biocatalyst for interesterification of soybean oil and lard. Food Chem 2016;194: 1283–1292. DOI: 10.1016/j.foodchem.2015.09.009.
- Silva RCD, Colleran HL, Ibrahim SA. Milk fat globule membrane in infant nutrition: A dairy industry perspective. J Dairy Res 2021; 88(1):105–116. DOI: 10.1017/S0022029921000224.
- 63. Knudsen J, Neergaard TB, Gaigg B, et al. Role of acyl-CoA binding protein in acyl-CoA metabolism and acyl-CoA-mediated cell signaling. J Nutr 2000;130(2S Suppl):294S–298S. DOI: 10.1093/jn/130.2.294S.
- 64. Hamosh M, Scanlon JW, Ganot D, et al. Fat digestion in the newborn. Characterization of lipase in gastric aspirates of premature and term infants. J Clin Invest 1981;67(3):838–846. DOI: 10.1172/jci110101.

- 65. Hamosh M. The role of lingual lipase in neonatal fat digestion. Ciba Found Symp 1979;(70):69–98. DOI: 10.1002/9780470720530.ch5.
- 66. Hamosh M. Lingual and gastric lipases. Nutrition 1990;6(6):421–428. PMID: 2134569.
- 67. Hamosh M. A review. Fat digestion in the newborn: Role of lingual lipase and preduodenal digestion. Pediatr Res 1979;13(5 Pt 1): 615–622. DOI: 10.1203/00006450-197905000-00008.
- 68. Olivecrona T, Hernell O. Human milk lipases and their possible role in fat digestion. Padiatr Padol 1976;11(4):600–604. PMID: 980524.
- 69. Wardell JM, Wright AJ, Bardsley WG, et al. Bile salt-stimulated lipase and esterase activity in human milk after collection, storage, and heating: Nutritional implications. Pediatr Res 1984;18(4):382–386. DOI: 10.1203/00006450-198404000-00017.
- Mazzocchi A, D'Oria V, De Cosmi V, et al. The role of lipids in human milk and infant formulae. Nutrients 2018;10(5):567. DOI: 10.3390/ nu10050567.
- Los-Rycharska E, Kieraszewicz Z, Czerwionka-Szaflarska M. Medium chain triglycerides (MCT) formulas in paediatric and allergological practice. Prz Gastroenterol 2016;11(4):226–231. DOI: 10.5114/pg. 2016.61374.
- 72. Burge K, Vieira F, Eckert J, et al. Lipid composition, digestion, and absorption differences among neonatal feeding strategies: Potential implications for intestinal inflammation in preterm infants. Nutrients 2021;13(2):550. DOI: 10.3390/nu13020550.
- 73. Delplanque B, Gibson R, Koletzko B, et al. Lipid quality in infant nutrition: Current knowledge and future opportunities. J Pediatr Gastroenterol Nutr 2015;61(1):8–17. DOI: 10.1097/MPG.000000000 000818.
- 74. Houten SM, Violante S, Ventura FV, et al. The biochemistry and physiology of mitochondrial fatty acid beta-oxidation and its genetic disorders. Annu Rev Physiol 2016;78:23–44. DOI: 10.1146/annurev-physiol-021115-105045.
- 75. Wang Y, Liu Z, Han Y, et al. Medium chain triglycerides enhances exercise endurance through the increased mitochondrial biogenesis and metabolism. PLoS One 2018;13(2):e0191182. DOI: 10.1371/journal. pone.0191182.
- Contarini G, Povolo M. Phospholipids in milk fat: Composition, biological and technological significance, and analytical strategies. Int J Mol Sci 2013;14(2):2808–2831. DOI: 10.3390/ijms14022808.
- 77. NC-IUBMB BNCola. On the nomenclature of fatty acids. 2022. https://iubmb.qmul.ac.uk/newsletter/2020.html.
- Nelson RH, Mundi MS, Vlazny DT, et al. Kinetics of saturated, monounsaturated, and polyunsaturated fatty acids in humans. Diabetes 2013;62(3):783–788. DOI: 10.2337/db12-0367.
- 79. German JB, Dillard CJ. Saturated fats: A perspective from lactation and milk composition. Lipids 2010;45(10):915–923. DOI: 10.1007/ s11745-010-3445-9.
- 80. He X, McClorry S, Hernell O, et al. Digestion of human milk fat in healthy infants. Nutr Res 2020;83:15–29. DOI: 10.1016/j.nutres.2020.08.002.
- Page KA, Williamson A, Yu N, et al. Medium-chain fatty acids improve cognitive function in intensively treated type 1 diabetic patients and support in vitro synaptic transmission during acute hypoglycemia. Diabetes 2009;58(5):1237–1244. DOI: 10.2337/db08-1557.
- 82. Yuan T, Wang L, Jin J, et al. Role medium-chain fatty acids in the lipid metabolism of infants. Front Nutr 2022;9:804880. DOI: 10.3389/ fnut.2022.804880.
- 83. Mu H, Hoy CE. Effects of different medium-chain fatty acids on intestinal absorption of structured triacylglycerols. Lipids 2000;35(1):83–89. DOI: 10.1007/s11745-000-0498-x.
- Huang CB, Alimova Y, Myers TM, et al. Short- and medium-chain fatty acids exhibit antimicrobial activity for oral microorganisms. Arch Oral Biol 2011;56(7):650–654. DOI: 10.1016/j.archoralbio.2011.01.011.
- Resh MD. Fatty acylation of proteins: The long and the short of it. Prog Lipid Res 2016;63:120–131. DOI: 10.1016/j.plipres.2016.05.002.
- 86. Cockshutt AM, Absolom DR, Possmayer F. The role of palmitic acid in pulmonary surfactant: Enhancement of surface activity and



prevention of inhibition by blood proteins. Biochim Biophys Acta 1991;1085(2):248–256. DOI: 10.1016/0005-2760(91)90101-m.

- Nguyen MTT, Kim J, Seo N, et al. Comprehensive analysis of fatty acids in human milk of four Asian countries. J Dairy Sci 2021;104(6): 6496–6507. DOI: 10.3168/jds.2020-18184.
- van de Vossenberg JL, Joblin KN. Biohydrogenation of C18 unsaturated fatty acids to stearic acid by a strain of Butyrivibrio hungatei from the bovine rumen. Lett Appl Microbiol 2003;37(5): 424–428. DOI: 10.1046/j.1472-765x.2003.01421.x.
- 89. Mancini A, Imperlini E, Nigro E, et al. Biological and nutritional properties of palm oil and palmitic acid: Effects on health. Molecules 2015;20(9):17339–17361. DOI: 10.3390/molecules200917339.
- Hernell O, Blackberg L. Digestion of human milk lipids: Physiologic significance of sn-2 monoacylglycerol hydrolysis by bile saltstimulated lipase. Pediatr Res 1982;16(10):882–885. DOI: 10.1203/ 00006450-198210000-00016.
- 91. Litmanovitz I, Davidson K, Eliakim A, et al. High beta-palmitate formula and bone strength in term infants: A randomized, doubleblind, controlled trial. Calcif Tissue Int 2013;92(1):35–41. DOI: 10.1007/ s00223-012-9664-8.
- 92. Kennedy K, Fewtrell MS, Morley R, et al. Double-blind, randomized trial of a synthetic triacylglycerol in formula-fed term infants: Effects on stool biochemistry, stool characteristics, and bone mineralization. Am J Clin Nutr 1999;70(5):920–927. DOI: 10.1093/ajcn/70.5.920.
- Gardner AS, Rahman IA, Lai CT, et al. Changes in fatty acid composition of human milk in response to cold-like symptoms in the lactating mother and infant. Nutrients 2017;9(9):1034. DOI: 10.3390/ nu9091034.
- 94. Siziba LP, Lorenz L, Brenner H, et al. Changes in human milk fatty acid composition and maternal lifestyle-related factors over a decade: A comparison between the two Ulm Birth Cohort Studies. Br J Nutr 2021;126(2):228–235. DOI: 10.1017/S0007114520004006.
- Miliku K, Duan QL, Moraes TJ, et al. Human milk fatty acid composition is associated with dietary, genetic, sociodemographic, and environmental factors in the CHILD cohort study. Am J Clin Nutr 2019; 110(6):1370–1383. DOI: 10.1093/ajcn/nqz229.
- Arnould VM, Reding R, Bormann J, et al. Predictions of daily milk and fat yields, major groups of fatty acids, and C18:1 cis-9 from single milking data without a milking interval. Animals (Basel) 2015;5(3):643–661. DOI: 10.3390/ani5030377.
- 97. Tyson J, Burchfield J, Sentance F, et al. Adaptation of feeding to a low fat yield in breast milk. Pediatrics 1992;89(2):215–220. PMID: 1734387.
- Mendonca MA, Araujo WMC, Borgo LA, et al. Lipid profile of different infant formulas for infants. PLoS One 2017;12(6):e0177812. DOI: 10.1371/journal.pone.0177812.
- Bobinski R, Bobinska J. Fatty acids of human milk A review. Int J Vitam Nutr Res 2022;92(3–4):280–291. DOI: 10.1024/0300-9831/ a000651.
- 100. Di Maso M, Bravi F, Ferraroni M, et al. Adherence to mediterranean diet of breastfeeding mothers and fatty acids composition of their human milk: Results from the Italian MEDIDIET study. Front Nutr 2022;9:891376. DOI: 10.3389/fnut.2022.891376.
- 101. Sanchez-Hernandez S, Esteban-Munoz A, Gimenez-Martinez R, et al. A comparison of changes in the fatty acid profile of human milk of Spanish lactating women during the first month of lactation using gas chromatography-mass spectrometry. A comparison with infant formulas. Nutrients 2019;11(12):3055. DOI: 10.3390/nu11123055.
- 102. Piccinin E, Cariello M, De Santis S, et al. Role of oleic acid in the gutliver axis: From diet to the regulation of its synthesis via stearoyl-CoA desaturase 1 (SCD1). Nutrients 2019;11(10). DOI: 10.3390/nu11102283.
- Samuel TM, Zhou Q, Giuffrida F, et al. Nutritional and non-nutritional composition of human milk is modulated by maternal, infant, and methodological factors. Front Nutr 2020;7:576133. DOI: 10.3389/ fnut.2020.576133.
- 104. Sioen I, van Lieshout L, Eilander A, et al. Systematic review on N-3 and N-6 polyunsaturated fatty acid intake in European countries in light of the current recommendations – Focus on specific population groups. Ann Nutr Metab 2017;70(1):39–50. DOI: 10.1159/000456723.

- Lopez-Lopez A, Lopez-Sabater MC, Campoy-Folgoso C, et al. Fatty acid and sn-2 fatty acid composition in human milk from Granada (Spain) and in infant formulas. Eur J Clin Nutr 2002;56(12):1242–1254. DOI: 10.1038/sj.ejcn.1601470.
- 106. Willatts P, Forsyth JS. The role of long-chain polyunsaturated fatty acids in infant cognitive development. Prostaglandins Leukot Essent Fatty Acids 2000;63(1–2):95–100. DOI: 10.1054/plef.2000.0198.
- 107. Abedi E, Sahari MA. Long-chain polyunsaturated fatty acid sources and evaluation of their nutritional and functional properties. Food Sci Nutr 2014;2(5):443–463. DOI: 10.1002/fsn3.121.
- Carlson SE, Colombo J. Docosahexaenoic acid and arachidonic acid nutrition in early development. Adv Pediatr 2016;63(1):453–471. DOI: 10.1016/j.yapd.2016.04.011.
- 109. Smith SL, Rouse CA. Docosahexaenoic acid and the preterm infant. Matern Health Neonatol Perinatol 2017;3(1):22. DOI: 10.1186/s40748-017-0061-1.
- 110. Heath RJ, Klevebro S, Wood TR. Maternal and neonatal polyunsaturated fatty acid intake and risk of neurodevelopmental impairment in premature infants. Int J Mol Sci 2022;23(2). DOI: 10.3390/ijms23020700.
- 111. Stark KD, Van Elswyk ME, Higgins MR, et al. Global survey of the omega-3 fatty acids, docosahexaenoic acid and eicosapentaenoic acid in the blood stream of healthy adults. Prog Lipid Res 2016;63: 132–152. DOI: 10.1016/j.plipres.2016.05.001.
- 112. Conway MC, McSorley EM, Mulhern MS, et al. The influence of fish consumption on serum n-3 polyunsaturated fatty acid (PUFA) concentrations in women of childbearing age: A randomised controlled trial (the iFish Study). Eur J Nutr 2021;60(3):1415–1427. DOI: 10.1007/s00394-020-02326-w.
- Gibson RA, Makrides M. Long-chain polyunsaturated fatty acids in breast milk: Are they essential? Adv Exp Med Biol 2001;501:375–383. DOI: 10.1007/978-1-4615-1371-1_46.
- 114. Bzikowska-Jura A, Czerwonogrodzka-Senczyna A, Jasinska-Melon E, et al. The concentration of omega-3 fatty acids in human milk is related to their habitual but not current intake. Nutrients 2019; 11(7):1585. DOI: 10.3390/nu11071585.
- 115. Floris LM, Stahl B, Abrahamse-Berkeveld M, et al. Human milk fatty acid profile across lactational stages after term and preterm delivery: A pooled data analysis. Prostaglandins Leukot Essent Fatty Acids 2020;156:102023. DOI: 10.1016/j.plefa.2019. 102023.
- Uauy R, Hoffman DR, Peirano P, et al. Essential fatty acids in visual and brain development. Lipids 2001;36(9):885–895. DOI: 10.1007/ s11745-001-0798-1.
- Agostoni C, Berni Canani R, Fairweather-Tait S. Scientific opinion on the essential composition of infant and follow-on formulae. EFSA J 2014;12(4):3760. DOI: 10.2903/j.efsa.2014.3760.
- 118. Le HD, Meisel JA, de Meijer VE, et al. The essentiality of arachidonic acid and docosahexaenoic acid. Prostaglandins Leukot Essent Fatty Acids 2009;81(2–3):165–170. DOI: 10.1016/j.plefa.2009.05.020.
- 119. Sabel KG, Lundqvist-Persson C, Bona E, et al. Fatty acid patterns early after premature birth, simultaneously analysed in mothers' food, breast milk and serum phospholipids of mothers and infants. Lipids Health Dis 2009;8:20. DOI: 10.1186/1476-511X-8-20.
- Demmelmair H, Koletzko B. Perinatal polyunsaturated fatty acid status and obesity risk. Nutrients 2021;13(11):3882. DOI: 10.3390/ nu13113882.
- 121. Pluymen LPM, Dalmeijer GW, Smit HA, et al. Long-chain polyunsaturated fatty acids in infant formula and cardiovascular markers in childhood. Matern Child Nutr 2018;14(2):e12523. DOI: 10.1111/mcn.12523.
- 122. Bopp M, Lovelady C, Hunter C, et al. Maternal diet and exercise: Effects on long-chain polyunsaturated fatty acid concentrations in breast milk. J Am Diet Assoc 2005;105(7):1098–1103. DOI: 10.1016/ j.jada.2005.04.004.
- 123. Duvall MG, Levy BD. DHA- and EPA-derived resolvins, protectins, and maresins in airway inflammation. Eur J Pharmacol 2016;785:144–155. DOI: 10.1016/j.ejphar.2015.11.001.

- 124. Yilmaz JL, Lim ZL, Beganovic M, et al. Determination of substrate preferences for desaturases and elongases for production of docosahexaenoic acid from oleic acid in engineered canola. Lipids 2017;52(3):207–222. DOI: 10.1007/s11745-017-4235-4.
- 125. Roszer T. Co-evolution of breast milk lipid signaling and thermogenic adipose tissue. Biomolecules 2021;11(11):1705. DOI: 10.3390/biom 11111705.
- 126. Smiddy MA, Huppertz T, van Ruth SM. Triacylglycerol and melting profiles of milk fat from several species. Int Dairy J 2012;24(2):64–69. DOI: 10.1016/j.idairyj.2011.07.001.
- 127. Rodriguez-Cruz M, Tovar AR, Palacios-Gonzalez B, et al. Synthesis of long-chain polyunsaturated fatty acids in lactating mammary gland: Role of Delta5 and Delta6 desaturases, SREBP-1, PPARalpha, and PGC-1. J Lipid Res 2006;47(3):553–560. DOI: 10.1194/jlr.M500407-JLR200.
- Suburu J, Gu Z, Chen H, et al. Fatty acid metabolism: Implications for diet, genetic variation, and disease. Food Biosci 2013;4:1–12. DOI: 10.1016/j.fbio.2013.07.003.
- 129. Salem N, Jr., Van Dael P. Arachidonic acid in human milk. Nutrients 2020;12(3):626. DOI: 10.3390/nu12030626.
- 130. van Wezel-Meijler G, van der Knaap MS, Huisman J, et al. Dietary supplementation of long-chain polyunsaturated fatty acids in preterm infants: Effects on cerebral maturation. Acta Paediatr 2002;91(9):942–950. DOI: 10.1080/080352502760272632.
- Lauritzen L, Brambilla P, Mazzocchi A, et al. DHA effects in brain development and function. Nutrients 2016;8(1):6. DOI: 10.3390/ nu8010006.
- 132. Zarate R, El Jaber-Vazdekis N, Tejera N, et al. Significance of long chain polyunsaturated fatty acids in human health. Clin Transl Med 2017;6(1):25. DOI: 10.1186/s40169-017-0153-6.
- 133. Miles EA, Childs CE, Calder PC. Long-chain polyunsaturated fatty acids (LCPUFAs) and the developing immune system: A narrative review. Nutrients 2021;13(1):247. DOI: 10.3390/nu13010247.
- Czosnykowska-Lukacka M, Lis-Kuberka J, Krolak-Olejnik B, et al. Changes in human milk immunoglobulin profile during prolonged lactation. Front Pediatr 2020;8:428. DOI: 10.3389/fped.2020.00428.
- Longo N, Frigeni M, Pasquali M. Carnitine transport and fatty acid oxidation. Biochim Biophys Acta 2016;1863(10):2422–2435. DOI: 10.1016/j.bbamcr.2016.01.023.
- 136. Ramaswamy M, Anthony Skrinska V, Fayez Mitri R, et al. Diagnosis of carnitine deficiency in extremely preterm neonates related to parenteral nutrition: Two step newborn screening approach. Int J Neonatal Screen 2019;5(3):29. DOI: 10.3390/ijns5030029.
- 137. Van Aerde JE. In preterm infants, does the supplementation of carnitine to parenteral nutrition improve the following clinical outcomes: Growth, lipid metabolism and apneic spells?: Part B: Clinical commentary. Paediatr Child Health 2004;9(8):573. DOI: 10.1093/pch/9.8.573.
- Cairns PA, Stalker DJ. Carnitine supplementation of parenterally fed neonates. Cochrane Database Syst Rev 2000;2000(4):CD000950. DOI: 10.1002/14651858.CD000950.
- Yaron S, Shachar D, Abramas L, et al. Effect of high beta-palmitate content in infant formula on the intestinal microbiota of term infants. J Pediatr Gastroenterol Nutr 2013;56(4):376–381. DOI: 10.1097/ MPG.0b013e31827e1ee2.
- 140. Wu W, Zhao A, Liu B, et al. Neurodevelopmental outcomes and gut bifidobacteria in term infants fed an infant formula containing high sn-2 palmitate: A cluster randomized clinical trial. Nutrients 2021;13(2):693. DOI: 10.3390/nu13020693.
- Peng Y, Zhou T, Wang Q, et al. Fatty acid composition of diet, cord blood and breast milk in Chinese mothers with different dietary habits. Prostaglandins Leukot Essent Fatty Acids 2009;81(5–6): 325–330. DOI: 10.1016/j.plefa.2009.07.004.
- 142. Juber BA, Jackson KH, Johnson KB, et al. Breast milk DHA levels may increase after informing women: A community-based cohort study from South Dakota USA. Int Breastfeed J 2016;12:7. DOI: 10.1186/ s13006-016-0099-0.

- 143. van Goor SA, Schaafsma A, Erwich JJ, et al. Mildly abnormal general movement quality in infants is associated with higher Mead acid and lower arachidonic acid and shows a U-shaped relation with the DHA/ AA ratio. Prostaglandins Leukot Essent Fatty Acids 2010;82(1):15–20. DOI: 10.1016/j.plefa.2009.11.004.
- 144. Luxwolda MF, Kuipers RS, Sango WS, et al. A maternal erythrocyte DHA content of approximately 6 g% is the DHA status at which intrauterine DHA biomagnifications turns into bioattenuation and postnatal infant DHA equilibrium is reached. Eur J Nutr 2012; 51(6):665–675. DOI: 10.1007/s00394-011-0245-9.
- Weseler AR, Dirix CE, Bruins MJ, et al. Dietary arachidonic acid dosedependently increases the arachidonic acid concentration in human milk. J Nutr 2008;138(11):2190–2197. DOI: 10.3945/jn.108.089318.
- 146. Smit EN, Koopmann M, Boersma ER, et al. Effect of supplementation of arachidonic acid (AA) or a combination of AA plus docosahexaenoic acid on breastmilk fatty acid composition. Prostaglandins Leukot Essent Fatty Acids 2000;62(6):335–340. DOI: 10.1054/plef.2000.0163.
- 147. van Goor SA, Dijck-Brouwer DA, Erwich JJ, et al. The influence of supplemental docosahexaenoic and arachidonic acids during pregnancy and lactation on neurodevelopment at eighteen months. Prostaglandins Leukot Essent Fatty Acids 2011;84(5–6):139–146. DOI: 10.1016/j.plefa.2011.01.002.
- 148. De Roos B, Mavrommatis Y, Brouwer IA. Long-chain n-3 polyunsaturated fatty acids: New insights into mechanisms relating to inflammation and coronary heart disease. Br J Pharmacol 2009; 158(2):413–428. DOI: 10.1111/j.1476-5381.2009.00189.x.
- 149. Richard C, Calder PC. Docosahexaenoic acid. Adv Nutr 2016;7(6): 1139–1141. DOI: 10.3945/an.116.012963.
- Tanaka K, Farooqui AA, Siddiqi NJ, et al. Effects of docosahexaenoic acid on neurotransmission. Biomol Ther (Seoul) 2012;20(2):152–157. DOI: 10.4062/biomolther.2012.20.2.152.
- Jeffrey BG, Weisinger HS, Neuringer M, et al. The role of docosahexaenoic acid in retinal function. Lipids 2001;36(9):859–871. DOI: 10.1007/s11745-001-0796-3.
- 152. Peters BD, Voineskos AN, Szeszko PR, et al. Brain white matter development is associated with a human-specific haplotype increasing the synthesis of long chain fatty acids. J Neurosci 2014;34(18):6367–6376. DOI: 10.1523/JNEUROSCI.2818-13.2014.
- Koletzko B, Agostoni C, Carlson SE, et al. Long chain polyunsaturated fatty acids (LC-PUFA) and perinatal development. Acta Paediatr 2001;90(4):460–464. PMID: 11332943.
- 154. Granot E, Jakobovich E, Rabinowitz R, et al. DHAD supplementation during pregnancy and lactation affects infants' cellular but not humoral immune response. Mediators Inflamm 2011;2011:493925. DOI: 10.1155/2011/493925.
- 155. Jensen CL, Maude M, Anderson RE, et al. Effect of docosahexaenoic acid supplementation of lactating women on the fatty acid composition of breast milk lipids and maternal and infant plasma phospholipids. Am J Clin Nutr 2000;71(1 Suppl):292S–299S. DOI: 10.1093/ajcn/71.1.292s.
- 156. Sherry CL, Oliver JS, Marriage BJ. Docosahexaenoic acid supplementation in lactating women increases breast milk and plasma docosahexaenoic acid concentrations and alters infant omega 6:3 fatty acid ratio. Prostaglandins Leukot Essent Fatty Acids 2015;95: 63–69. DOI: 10.1016/j.plefa.2015.01.005.
- 157. Gibson RA, Neumann MA, Makrides M. Effect of increasing breast milk docosahexaenoic acid on plasma and erythrocyte phospholipid fatty acids and neural indices of exclusively breast fed infants. Eur J Clin Nutr 1997;51(9):578–584. DOI: 10.1038/sj.ejcn.1600446.
- Marc I, Plourde M, Lucas M, et al. Early docosahexaenoic acid supplementation of mothers during lactation leads to high plasma concentrations in very preterm infants. J Nutr 2011;141(2):231–236. DOI: 10.3945/jn.110.125880.
- Basak S, Mallick R, Duttaroy AK. Maternal docosahexaenoic acid status during pregnancy and its impact on infant neurodevelopment. Nutrients 2020;12(12):3615. DOI: 10.3390/nu12123615.



- 160. Khandelwal S, Kondal D, Chaudhry M, et al. Effect of maternal docosahexaenoic acid (DHA) supplementation on offspring neurodevelopment at 12 months in India: A randomized controlled trial. Nutrients 2020;12(10). DOI: 10.3390/nu12103041.
- Greenberg JA, Bell SJ, Ausdal WV. Omega-3 fatty acid supplementation during pregnancy. Rev Obstet Gynecol Fall 2008;1(4):162–169. PMCID: PMC2621042.
- 162. Salas Lorenzo I, Chisaguano Tonato AM, de la Garza Puentes A, et al. The effect of an infant formula supplemented with AA and DHA on fatty acid levels of infants with different FADS genotypes: The COGNIS study. Nutrients 2019;11(3):602. DOI: 10.3390/nu 11030602.
- Lien EL, Richard C, Hoffman DR. DHA and ARA addition to infant formula: Current status and future research directions. Prostaglandins Leukot Essent Fatty Acids 2018;128:26–40. DOI: 10.1016/ j.plefa.2017.09.005.
- 164. Gould JF, Colombo J, Collins CT, et al. Assessing whether early attention of very preterm infants can be improved by an omega-3 long-chain polyunsaturated fatty acid intervention: A follow-up of a randomised controlled trial. BMJ Open 2018;8(5):e020043. DOI: 10.1136/bmjopen-2017-020043.
- Makrides M, Gibson RA, McPhee AJ, et al. Neurodevelopmental outcomes of preterm infants fed high-dose docosahexaenoic acid: A randomized controlled trial. JAMA 2009;301(2):175–182. DOI: 10.1001/ jama.2008.945.
- Qawasmi A, Landeros-Weisenberger A, Bloch MH. Meta-analysis of LCPUFA supplementation of infant formula and visual acuity. Pediatrics 2013;131(1):e262–e272. DOI: 10.1542/peds.2012-0517.
- 167. Pietrzak-Fiecko R, Kamelska-Sadowska AM. The comparison of nutritional value of human milk with other mammals' milk. Nutrients 2020;12(5):1404. DOI: 10.3390/nu12051404.
- Li Y, Gao D, Chen L, et al. The association between breastfeeding duration and lipid profile among children and adolescents. Nutrients 2021;13(8):2728. DOI: 10.3390/nu13082728.
- 169. Parikh NI, Hwang SJ, Ingelsson E, et al. Breastfeeding in infancy and adult cardiovascular disease risk factors. Am J Med 2009;122(7): 656–663.e1. DOI: 10.1016/j.amjmed.2008.11.034.
- 170. Suganuma H, Collins CT, McPhee AJ, et al. Effect of parenteral lipid emulsion on preterm infant PUFAs and their downstream metabolites. Prostaglandins Leukot Essent Fatty Acids 2021;164:102217. DOI: 10.1016/j.plefa.2020.102217.
- 171. Hellstrom A, Hellstrom W, Hellgren G, et al. Docosahexaenoic acid and arachidonic acid levels are associated with early systemic inflammation in extremely preterm infants. Nutrients 2020;12(7):1996. DOI: 10.3390/nu12071996.
- 172. Valentine CJ, Dingess KA, Kleiman J, et al. A randomized trial of maternal docosahexaenoic acid supplementation to reduce inflammation in extremely preterm infants. J Pediatr Gastroenterol Nutr 2019;69(3):388–392. DOI: 10.1097/MPG.00000000002375.
- 173. Ulu A, Burr A, Heires AJ, et al. A high docosahexaenoic acid diet alters lung inflammation and recovery following repetitive exposure to aqueous organic dust extracts. J Nutr Biochem 2021;97:108797. DOI: 10.1016/j.jnutbio.2021.108797.
- Sodhi CP, Fulton WB, Good M, et al. Fat composition in infant formula contributes to the severity of necrotising enterocolitis. Br J Nutr 2018;120(6):665–680. DOI: 10.1017/S0007114518001836.
- 175. Kim HY, Spector AA. N-Docosahexaenoylethanolamine: A neurotrophic and neuroprotective metabolite of docosahexaenoic acid. Mol Aspects Med 2018;64:34–44. DOI: 10.1016/j.mam.2018. 03.004.
- 176. Kim HY, Huang BX, Spector AA. Molecular and signaling mechanisms for docosahexaenoic acid-derived neurodevelopment and neuroprotection. Int J Mol Sci 2022;23(9):4635. DOI: 10.3390/ijms 23094635.
- 177. Akerele OA, Cheema SK. Maternal diet high in Omega-3 fatty acids upregulate genes involved in neurotrophin signalling in fetal brain during pregnancy in C57BL/6 mice. Neurochem Int 2020;138:104778. DOI: 10.1016/j.neuint.2020.104778.

- Ruiz-Lopez N, Usher S, Sayanova OV, et al. Modifying the lipid content and composition of plant seeds: engineering the production of LC-PUFA. Appl Microbiol Biotechnol 2015;99(1):143–154. DOI: 10.1007/ s00253-014-6217-2.
- Brenna JT, Kothapalli KS, Park WJ. Alternative transcripts of fatty acid desaturase (FADS) genes. Prostaglandins Leukot Essent Fatty Acids 2010;82(4–6):281–285. DOI: 10.1016/j.plefa.2010.02.011.
- 180. Park HG, Park WJ, Kothapalli KS, et al. The fatty acid desaturase 2 (FADS2) gene product catalyzes Delta4 desaturation to yield n-3 docosahexaenoic acid and n-6 docosapentaenoic acid in human cells. FASEB J 2015;29(9):3911–3919. DOI: 10.1096/fj.15-271783.
- Park WJ, Kothapalli KS, Reardon HT, et al. A novel FADS1 isoform potentiates FADS2-mediated production of eicosanoid precursor fatty acids. J Lipid Res 2012;53(8):1502–1512. DOI: 10.1194/jlr.M025312.
- 182. Emery JA, Hermon K, Hamid NK, et al. Delta-6 Desaturase substrate competition: dietary linoleic acid (18:2n-6) has only trivial effects on alpha-linolenic acid (18:3n-3) bioconversion in the teleost rainbow trout. PLoS One 2013;8(2):e57463. DOI: 10.1371/journal.pone.0057463.
- 183. Rayyan M, Devlieger H, Jochum F, et al. Short-term use of parenteral nutrition with a lipid emulsion containing a mixture of soybean oil, olive oil, medium-chain triglycerides, and fish oil: a randomized double-blind study in preterm infants. JPEN J Parenter Enteral Nutr 2012;36(1 Suppl):81S–94S. DOI: 10.1177/0148607111424411.
- Raman M, Almutairdi A, Mulesa L, et al. Parenteral nutrition and lipids. Nutrients 2017;9(4):388. DOI: 10.3390/nu9040388.
- Nilsson AK, Lofqvist C, Najm S, et al. Influence of human milk and parenteral lipid emulsions on serum fatty acid profiles in extremely preterm infants. JPEN J Parenter Enteral Nutr 2019;43(1):152–161. DOI: 10.1002/jpen.1172.
- Klek S. Omega-3 fatty acids in modern parenteral nutrition: A review of the current evidence. J Clin Med 2016;5(3). DOI: 10.3390/ jcm5030034.
- Guthrie G, Burrin D. Impact of parenteral lipid emulsion components on cholestatic liver disease in neonates. Nutrients 2021;13(2). DOI: 10.3390/nu13020508.
- Cleminson J, McGuire W, Embleton N. Commentary on "Lipid Emulsions for Parenterally Fed Preterm Infants". Neonatology 2021;118(1):1–4. DOI: 10.1159/000512679.
- 189. Kapoor V, Glover R, Malviya MN. Alternative lipid emulsions versus pure soy oil based lipid emulsions for parenterally fed preterm infants. Cochrane Database Syst Rev 2015;2015(12):CD009172. DOI: 10.1002/14651858.CD009172.pub2.
- 190. Kapoor V, Malviya MN, Soll R. Lipid emulsions for parenterally fed term and late preterm infants. Cochrane Database Syst Rev 2019;6(6):CD013171. DOI: 10.1002/14651858.CD013171.pub2.
- 191. Lofqvist CA, Najm S, Hellgren G, et al. Association of retinopathy of prematurity with low levels of arachidonic acid: A secondary analysis of a randomized clinical trial. JAMA Ophthalmol 2018;136(3):271–277. DOI: 10.1001/jamaophthalmol.2017.6658.
- 192. Hellstrom A, Pivodic A, Granse L, et al. Association of docosahexaenoic acid and arachidonic acid serum levels with retinopathy of prematurity in preterm infants. JAMA Netw Open 2021;4(10):e2128771. DOI: 10.1001/jamanetworkopen.2021.28771.
- Moon K, Rao SC, Schulzke SM, et al. Longchain polyunsaturated fatty acid supplementation in preterm infants. Cochrane Database Syst Rev 2016;12(12):CD000375. DOI: 10.1002/14651858.CD000375.pub5.
- Elliott E, Hanson CK, Anderson-Berry AL, et al. The role of specialized pro-resolving mediators in maternal-fetal health. Prostaglandins Leukot Essent Fatty Acids 2017;126:98–104. DOI: 10.1016/j.plefa.2017. 09.017.
- Lapillonne A. Enteral and parenteral lipid requirements of preterm infants. World Rev Nutr Diet 2014;110:82–98. DOI: 10.1159/000358460.
- Middleton P, Gomersall JC, Gould JF, et al. Omega-3 fatty acid addition during pregnancy. Cochrane Database Syst Rev 2018;11:CD003402. DOI: 10.1002/14651858.CD003402.pub3.
- Atkins D, Best D, Briss PA, et al. Grading quality of evidence and strength of recommendations. BMJ 2004;328(7454):1490. DOI: 10.1136/bmj.328.7454.1490.

- 198. Boquien CY. Human milk: An ideal food for nutrition of preterm newborn. Front Pediatr 2018;6:295. DOI: 10.3389/fped.2018.00295.
- Underwood MA. Human milk for the premature infant. Pediatr Clin North Am 2013;60(1):189–207. DOI: 10.1016/j.pcl.2012.09.008.
- Wallenborn JT, Valera CB, Kounnavong S, et al. Urban-rural gaps in breastfeeding practices: Evidence from Lao People's Democratic Republic. Int J Public Health 2021;66:1604062. DOI: 10.3389/ijph. 2021.1604062.
- 201. Abegunde D, Hutchinson P, Anaba U, et al. Socioeconomic inequality in exclusive breastfeeding behavior and ideation factors for social behavioral change in three north-western Nigerian states: A crosssectional study. Int J Equity Health 2021;20(1):172. DOI: 10.1186/ s12939-021-01504-4.
- 202. Infant and young child feeding. 2022, 2021. https://www.who.int/ news-room/fact-sheets/detail/infant-and-young-child-feeding.
- 203. Breastfeeding. https://www.thelancet.com/series/breastfeeding.
- Victora CG, Bahl R, Barros AJD. Breastfeeding in the 21st century: Epidemiology, mechanisms, and lifelong effect. Lancet 2016; 387(10017):475–490. DOI: 10.1016/S0140-6736(15)01024-7.
- Rollins NC, Bhandari N, Hajeebhoy N. Why invest, and what it will take to improve breastfeeding practices? Lancet 2016;387(10017):491–504. DOI: 10.1016/S0140-6736(15)01044-2.

- 206. Jones KM, Power ML, Queenan JT, et al. Racial and ethnic disparities in breastfeeding. Breastfeed Med 2015;10(4):186–196. DOI: 10.1089/ bfm.2014.0152.
- Heck KE, Braveman P, Cubbin C, et al. Socioeconomic status and breastfeeding initiation among California mothers. Public Health Rep 2006;121(1):51–59. DOI: 10.1177/003335490612100111.
- 208. Pierro J, Abulaimoun B, Roth P, et al. Factors associated with supplemental formula feeding of breastfeeding infants during postpartum hospital stay. Breastfeed Med 2016;11:196–202. DOI: 10.1089/bfm.2015.0091.
- 209. Salminen S, Stahl B, Vinderola G, et al. Infant formula supplemented with biotics: Current knowledge and future perspectives. Nutrients 2020;12(7):1952. DOI: 10.3390/nu12071952.
- 210. Carlson SE. Early determinants of development: A lipid perspective. Am J Clin Nutr 2009;89(5):1523S–1529S. DOI: 10.3945/ajcn.2009.27113G.
- 211. Ramiro-Cortijo D, Singh P, Liu Y, et al. Breast milk lipids and fatty acids in regulating neonatal intestinal development and protecting against intestinal injury. Nutrients 2020;12(2):534. DOI: 10.3390/nu12020534.
- Bayley TM, Alasmi M, Thorkelson T, et al. Influence of formula versus breast milk on cholesterol synthesis rates in four-month-old infants. Pediatr Res 1998;44(1):60–67. DOI: 10.1203/00006450-199807000-00010.